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ISOLATION OF PTEROPODIC ACID FROM MALAYSIAN Uncaria lanosa VAR. ferrea BY USING LIQUID CHROMATOGRAPHY MASS SPECTROMETRY DEREPLICATION APPROACH

(Pemencilan Asid Pteropodik daripada *Uncaria lanosa* var. *ferrea* Malaysia dengan Pendekatan Direplikasi Kromatografi Cecair Spektrometri Jisim)

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Abstract

Previous chemical profiling on Malaysian Uncaria lanosa var. ferrea using liquid chromatography mass spectrometry (LCMS) had successfully identified three alkaloids namely pteropodine, isopteropodine, and isopteropodic acid along with a flavonoid known as rutin. Continuing our interest on this species, a dereplication strategy was applied to target unknown peak from the plant material. This had successfully led to the isolation of a new alkaloid. The molecular structure of the isolated new alkaloid was elucidated by using various spectroscopic techniques and was recognized as an acidic derivative of the alpha-beta unsaturated carbonyl methyl ester of pteropodine, one of the previously identified alkaloids from the plant. The new alkaloid was named as pteropodic acid. This study demonstrates that LCMS dereplication strategy will not only lead to an increased possibility of discovery of new compounds but also will save time, energy, and resources.

Keywords: Uncaria, liquid chromatography mass spectrometry, dereplication, alkaloid

Abstrak

Profil kimia sebelumnya oleh Uncaria lanosa var. ferrea Malaysia melalui kromatografi cecair spektrometer jisim (LCMS) berjaya mengenal pasti tiga alkaloid iaitu pteropodin, isopteropodin, dan asid isopteropodik bersama dengan flavonoid yang dikenali sebagai rutin. Meneruskan minat kami terhadap spesies ini, strategi direplikasi digunakan untuk menyasarkan puncak yang tidak diketahui dari bahan tumbuhan yang sama dan ini berjaya memencilkan alkaloid baru. Struktur molekul alkaloid baru yang dipencilkan telah dipastikan dengan menggunakan pelbagai teknik spektroskopi dan ditentukan sebagai molekul berasid terbitan karbonil metil ester tak tepu alpha-beta dari pteropodin, iaitu salah satu alkaloid yang dikenal pasti sebelumnya dalam tumbuhan yang sama. Oleh itu alkaloid baru ini dinamakan sebagai asid pteropodik. Dapatan kajian ini menunjukkan bahawa strategi direplikasi LCMS tidak hanya akan meningkatkan kemungkinan dalam menemui sebatian baru tetapi juga mampu menjimatkan waktu, tenaga, dan sumber bahan.

Kata kunci: Uncaria, kromatografi cecair spektrometer jisim, direplikasi, alkaloid

Introduction

Genus Uncaria, a major part of Rubiaceae family, comprising 34 species has been used to treat various diseases worldwide [1]. To date, over 400 compounds identified in *Uncaria* have been primarily alkaloids, terpenoids, and flavonoids [2]. Uncaria lanosa Wall, locally known as "gegambir paya" or "gegambir hitam", is one of 14 species of Uncaria available in Malaysia [3]. The plant is reported to be used as an infusion for intestine inflammation and a decoction for cleaning wounds and ulcers. According to Turner [4], there are 11 varieties of Uncaria lanosa Wall. These are Uncaria lanosa var. appendiculata (Benth.) Ridsdale, *Uncaria lanosa* var. *ferrea* (Blume) Ridsdale, Uncaria lanosa f. ferrea (Blume) Ridsdale, Uncaria lanosa var. glabrata (Blume) Ridsdale, Uncaria lanosa f. glabrescens (Merr. & Perry) Ridsdale, Uncaria lanosa f. gynogumna Ridsdale, Uncaria lanosa var. korrensis (Kaneh.) Ridsdale, Uncaria lanosa f. philippinensis (Elmer) Ridsdale, Uncaria lanosa f. setiloba (Benth.) Ridsdale, Uncaria lanosa f. sumatrana Ridsdale, and Uncaria lanosa var. toppingii (Merr.) Ridsdale.

In the last two decades, the mass spectrometry technique had undergone a rapid development, allowing analysis of natural products with differentiated physicochemical properties. This is due to the instrument versatility and sensitivity that could be applied to structural metabolite mixtures, of plant characterization particularly for identifying minor components [5]. Liquid chromatography-mass spectrometry (LCMS)based fingerprinting is one of the methods that would be able to determine the genetic diversity and the authentication of a plant species. In addition, an application of a strategic method such as dereplication would further facilitate and accelerate the metabolite identification, hence search for a novel active compound. The dereplication method requires to compare molecular chromatographic and spectroscopic

data characteristics possibly obtained from an online database or an in-house library [6,7].

Our previous chemical profiling on the dichloromethane (DCM) stem extract of Malaysian *Uncaria lanosa* var. *ferrea* through Orbitrap LCMS successfully identify three alkaloids namely pteropodine, isopteropodine, and isopteropodic acid along with a flavonoid known as rutin [8]. In the present work, an LCMS dereplication strategy was applied to target for unknown peak from the same plant material, and this had led to the isolation of a new alkaloid. The dereplication process which had led to the isolation and the structural identification of the new alkaloid are discussed here.

Materials and Methods

Chemicals and raw materials

Stems of *U. lanosa var farrea* (Blume) Ridsd. were collected from Hutan Simpan Pasir Raja, Dungun Terengganu, Malaysia. This plant was identified by Mr. Ahmad Zainuddin Ibrahim from Universiti Kebangsaan Malaysia and the specimen voucher (HTBP4324) was deposited at the Herbarium Taman Botani Putrajaya, Malaysia. Ultrapure water was obtained from a PURELAB Ultra Laboratory Water Purification Systems while HPLC grade acetonitrile (MeCN) and methanol (MeOH) were purchased from Merck. Solvents used for fractionation and isolation were of industrial grade and were distilled prior to use.

Sample preparation

Dried and ground stems of *Uncaria lanosa* (1.38 kg) was extracted through 3-day maceration with methanol. The MeOH was removed until complete dryness, and crude MeOH extract (88.50 g) was further triturated using two types of solvents; non-polar solvent (hexane) and medium- polar solvent (DCM). Each mixture was filtered and evaporated under reduced pressure which produced 3 extracts; hexane (5.78 g), DCM (15.15 g) and MeOH (49.45 g). The DCM extract was subjected to an on open column chromatography [DCM/MeOH

(100:0 – 20:80)] with gradual increase in solvent polarity to yield seven potential fractions. 1 mg of the crude extract and fractions were weighed accurately and dissolved in 1 mL of MeOH: water containing 0.1% formic acid (30:70, v/v). Samples were filtered through a 0.22 µm PTFE filter into 2 ml screw cap vials prior to LCMS analysis.

High performance liquid chromatography mass spectrometry conditions

The optimized methodology in this study is slightly different compared to previous paper [7]. Samples including extracts and 17 reference compounds (1 mg/mL) were separated using GL Sciences - Inertsil Column C-18 (250 mm x 4.6 mm, i.d., 5 μ m). The flow rate was set at 0.6 mL/min, the absorbance was set to 254 and 215 nm, and the injection volume was 5 μ L. After each injection the needle was washed with 100% MeCN. The mobile phase was a mixture of water (A) and MeCN (B) both containing 0.01% formic acid with gradient elution program of 5-55% B (0.00-30 min), 55-100% B (30-40 min), and 100% B (10-45 min).

HPLC-DAD-TOF MS was conducted on Agilent 1100 Series of high-performance liquid chromatography coupled with diode-array and time-of-flight mass detectors, electrospray ionization (ESI) source system, dual pump, autosampler and column. Nitrogen was used as the nebulizing and dissolving gas with a flow rate of 50 and 400 Lit/h, respectively. The optimum source of ESI conditions were: 3.5kV spray voltage, 25V cone voltage, 3V extractor, 0.2V rf lens, 100°C source temperature, and 350°C probe temperature. Data processing and library searching was performed using Qualitative Analysis on Mass Hunter Acquisition Data.

Isolation of pteropodic acid using recycling HPLC

One of the potential fractions, D4 (639.80 mg) was targeted for further isolation. The fraction was subjected to recycling HPLC using two column Jaigel- ODS- AP, SP- 120-15 (250 mm x 20 mm, i.d., 5 μ m) with guard column and GL Sciences- Inertsustain Column C-18 (250 mm x 20 mm, i.d., 5 μ m). The flow rate of the system was set at 4 mL/min, the absorbance at 254 nm, and the injection volume was 1 mL. The fraction was separated using an isocratic elution (20:80) of water and

MeCN both containing 0.01% formic acid. The new alkaloid, pteropodic acid (5.7 mg) was eluted in the first cycle at minutes 35.

Results and Discussion

Development of in-house database

The LC-ESI-MS profiles of previously isolated compounds from *Uncaria* species were added into inhouse database. This database was used as reference for the identification of compounds from LCMS data analysis of the crude extracts and fractions of *Uncaria* species for the isolation of targeted compounds. The LCMS profiles are shown in Table 1.

LCMS dereplication analysis of *Uncaria lanosa* extract

The HPLC-DAD-TOF-MS system was employed for the metabolite profiling and identification of the chromatographic peak of the DCM stem extract of U. lanosa var. ferrea. From the analysis, a positive mode affords higher sensitivity for most of the peaks compared to a negative mode. Therefore, a positive mode was chosen and applied in further analyses. By comparing the retention time and m/z value to the reference compounds in the in-house database, only four peaks were identified, including one flavonoid which was rutin (6), and three alkaloids which were pteropodine (8), isopterpodic acid (10), isopteropodine (13). As shown in Figure 1, four peaks were successfully dereplicated. These were peaks (6), (8), (10), and (13) with $[M+H]^+$ ion of m/z 611.1624, 369.2173, 355.1779 and 369.2250, respectively. A major unknown $[M+H]^+$ ion peak at m/z 355.2707 was targeted for further isolation using recycling HPLC.

The targeted compound was isolated as white amorphous powder and its mass spectrum gave an [M+H]⁺ peak at m/z 355.2707 which corresponded to the molecular formula $C_{20}H_{22}N_2O_4$. Its UV spectrum displayed absorption bands at 209.97 and 244.00 nm indicating that a hydroxyl substituent may be present. The IR spectrum revealed the presence of non-hydrogen-bonded OH (sharp peak) at 3421.96 cm⁻¹. Typical IR characteristics of heteroyohim B bine-type oxindole alkaloids absorption frequencies at 3114 cm⁻¹ (NH), 1622.02 cm⁻¹ (C=O acid), 1674 cm⁻¹ (C=O

amide), 1636 cm⁻¹ (C=C olefinic), 1472.8 cm⁻¹ (C=C aromatic) and 1191.42 cm⁻¹ (C-O cyclic ether) were also observed suggesting that the compound belonged to pentacyclic oxindole alkaloid. The ¹H NMR spectrum of this compound showed that four aromatic proton signals resonating at δ 7.29, δ 7.21, 7.07 and 6.88 confirming that the presence of aromatic ring is unsubstituted in the oxindole moiety. Proton resonances were also observed as a singlet at δ 7.50 belonging to an olefinic proton H-17 and a broad singlet at δ 8.56 attributed to a proton attached with nitrogen. A quartet and three-proton doublet resonances at δ 4.46 (J = 6.1 Hz) and 1.41 (J =6Hz) were also present corresponding to H-19 and H-18 respectively. There was no carbomethoxy (OCH₃) resonance which was typically observed in other POAs H¹ NMR spectrum [9, 10].

In the ^{13}C NMR spectrum, the presence of 20 carbons including a methyl group, an acid carbonyl and oxindole carbonyl resonances at δ 17.59, δ 170.40 and δ 181.92 respectively. The slightly higher ^{13}C NMR resonance value for the acid carbonyl (δ 170.40) compared to the ester carbonyl in the pteropodine system (δ 167.77) further confirmed the existence of a carboxylic acid moiety in the structure [11]. Therefore, based on these spectral data as well as comparisons with literature data, the compound was deduced to be pteropodic acid which is the acidic derivative of the alpha-beta unsaturated

carbonyl methyl ester of pteropodine, one of the previously identified alkaloids from the plant. The structure of the dereplicated and the isolated alkaloid is shown in Figure 2.

Physical and spectral data of isolated compound

Pteropodic acid: White amorphous powder, wt: 5.7 mg (MeOH), mp 268 - 271 °C. MS m/z: 355.2707 [M+H]⁺, $C_{20}H_{22}N_2O_4$; UV (MeOH) λ_{max} nm: 209.97, 244.00; IR (KBr) v_{max} cm⁻¹: 3421.96 (OH), 3114 (NH), 1622.02 (C=O acid), 1674 (C=O amide), 1636 (C=C olefinic), 1472.8 (C=C aromatic) and 1191.42 (C-O cyclic ether); ¹H NMR (MeOD, 600 MHz) δ ppm: 8.56 (1H, br s, N-H), 7.50 (1H, s, H-17), 7.29 (1H, d, J = 7.2 Hz, H-9), 7.21 (1H, ddd, J = 7.7, 1.0 Hz, H-11), 7.07 (1H, ddd, J =7.6, 0.8 Hz, H-10), 6.88 (1H, d, J = 7.7 Hz, H-12), 4.46 $(1H, q, J = 6.1 \text{ Hz}, H-19), 3.39 - 3.34 (2H, m, H-21\beta, H 5\beta$), 2.50 - 2.27 (5H, m, H-15, H-21α, H-6β, H-5α, H-3), 2.07-1.99 (1H, m, H-6 α), 1.70-1.64 (2H, br m, H- 14α , H-20), 1.63-1.57 (1H, m, H-14 β), 1.41 (3H, d, J =6 Hz, CH₃); ¹³C NMR (MeOD, 150 MHz) δ ppm: 181.92 (N-C=O), 170.40 (O-C=O), 154.60 (C-17), 141.50 (C-13), 133.30 (C-8), 127.70 (C-11), 122.79 (C-9), 122.27 (C-10), 110.39 (C-16), 109.09 (C-12), 74.11 (C-19), 71.85 (C-3), 56.42 (C-7), 54.59 (C-5), 53.03 (C-21), 38.11 (C-20), 33.56 (C-6), 31.16 (C-15), 29.55 (C-14), 17.59 (CH₃).

Table 1. Characterization of reference compounds by developed HPLC-DAD-TOF MS method

| No. | Compound (RT (min)) | Proposed Molecular Formula | Observed Mass [M+H] ⁺ | Calculated Mass [M+H] ⁺ | Observed Mass [M-H] ⁻ | Calculated Mass [M-H] ⁻ |
|-----|----------------------------------|----------------------------------|--|--|--|--|
| 1 | Formosaninol (10.23) | $C_{21}H_{26}N_2O_5$ | 387.2092 | 387.1920 | - | 385.1764 |
| 2 | Catechin (10.63) | $C_{15}H_{14}O_6$ | 291.1058 | 291.0868 | 289.0612 | 289.0712 |
| 3 | Isoformosaninol (11.65) | $C_{21}H_{26}N_2O_5$ | 387.2108 | 387.1920 | - | 385.1764 |
| 4 | (-) <i>Epi</i> -catechin (12.77) | $C_{15}H_{14}O_6$ | 291.1058 | 291.0868 | 289.0600 | 289.0712 |
| 5 | (-)Epi-afzelechin (15.15) | $C_{15}H_{14}O_5$ | 275.1117 | 275.0919 | 273.0661 | 273.0763 |
| 6 | Rutin (16.88) | $C_{27}H_{30}O_{16}$ | 611.1745 | 611.1612 | 609.1128 | 609.1456 |
| 7 | Scopoletin (17.49) | $C_{10}H_8O_4$ | 193.0726 | 193.0501 | - | 191.0345 |
| 8 | Pteropodine (17.64) | $C_{21}H_{24}N_2O_4$ | 369.1971 | 369.1814 | - | 367.1658 |
| 9 | Longiflorine (17.69) | $C_{11}H_{15}NO_4$ | 226.1280 | 226.1079 | - | 224.0923 |
| 10 | Isopteropodic acid (18.32) | $C_{20}H_{22}N_2O_4$ | 355.1841 | 355.1658 | - | 353.1502 |

| Table 1 (cont'd). Char | racterization of reference com | pounds by develor | oed HPLC-DAD-TOF MS method |
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|------------------------|--------------------------------|-------------------|----------------------------|

| No. | Compound (RT (min)) | Proposed Molecular Formula | Observed Mass [M+H] ⁺ | Calculated Mass [M+H] ⁺ | Observed Mass [M-H] ⁻ | Calculated Mass [M-H] |
|-----|--|----------------------------------|--|--|--|-----------------------------|
| 11 | Methyl paraben (21.72) | $C_8H_8O_3$ | 153.0777 | 153.0552 | 151.0387 | 151.0396 |
| 12 | Uncariechin (22.64) | $C_{18}H_{14}O_{6}$ | 327.1062 | 327.0868 | 325.0564 | 325.0712 |
| 13 | Isopterodine (23.67) | $C_{21}H_{24}N_2O_4$ | 369.1996 | 369.1814 | - | 367.1658 |
| 14 | Uncarine F (23.75) | $C_{21}H_{24}N_2O_4$ | 369.2063 | 369.1814 | 367.1625 | 367.1658 |
| 15 | Speciophylline (23.79) | $C_{21}H_{24}N_2O_4$ | 369.2126 | 369.1814 | 367.1634 | 367.1658 |
| 16 | Rauniticine- <i>allo</i> -oxindole B (23.83) | $C_{21}H_{24}N_2O_4$ | 369.2182 | 369.1814 | 367.1566 | 367.1658 |
| 17 | Quercetin (24.72) | $C_{15}H_{10}O_7$ | 303.0699 | 303.0505 | 301.0248 | 301.0349 |

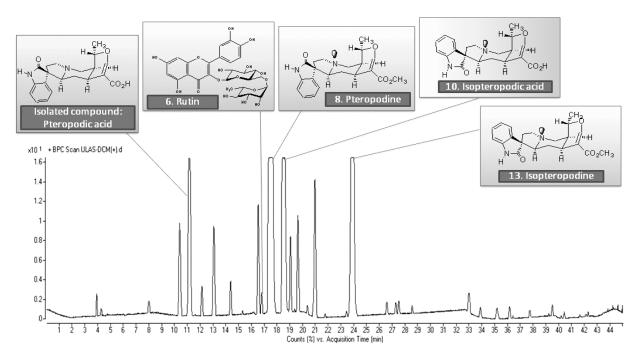


Figure 1. MS profile of *U. lanosa* DCM extract; the total ion current (TIC) in positive mode with structure of identified peaks; numbers of identified peak are according to the no. of reference compound in Table 1.

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Figure 2. Structure of isolated alkaloid from Uncaria lanosa var. ferrea identified as pteropodic acid

Conclusion

The proposed HPLC-DAD-TOF-MS dereplication strategy using in-house developed database was applied to identify known peaks and target unknown peak from *U. lanosa* var. *ferrea*. This has successfully led to the identification of one flavonoid and three alkaloids, and the isolation of a new alkaloid which was deduced as pteropodic acid. This study demonstrates that the LCMS dereplication strategy will not only increase the possibility discovery of new compounds but will save time, energy and resources by minimizing repetitive isolation of known compounds.

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References

- Ahmad, R., and Salim, F. (2015). Oxindole alkaloids of *Uncaria* (Rubiaceae, subfamily cinchonoideae): A review on its structure, properties, and bioactivities. In *Book Studies in Natural Products Chemistry*, 45: 486-525.
- Qin, N., Lu, X., Liu, Y., Qiao, Y., Qu, W., Feng, F. and Sun, H. (2021). Recent research progress of *Uncaria* spp. based on alkaloids: phytochemistry, pharmacology and structural chemistry. *European Journal of Medicinal Chemistry*, 210: 112960.
- 3. Burkill, I. H. (1966). A dictionary of the economic

- products of Malaysia Peninsula. 2nd edition. Kuala Lumpur: pp. 2236-2245.
- 4. Turner, I. M. (2018). A revised conspectus of *Uncaria* (Rubiaceae). *Journal of Plant Taxonomy and Geography*, 73(1): 9-21.
- 5. Alvarez-rivera, G., Ballesteros-vivas, D., Paradaalfonso, F., Ibanez, E., and Cifuentes, A. (2019). Recent applications of high resolution mass spectrometry for the characterization of plant natural products. *Trends in Analytical Chemistry*, 112: 87-101.
- Lone, S. A., Kushwaha, M., Wani, A., Kumar, A., Gupta, A. P., Hassan, Q. P., Chandra, S. and Gupta, S. (2017). Genetic diversity, LCMS based chemical fingerprinting and antioxidant activity of *Epimedium elatum* Morr & Decne. *Journal of Applied Research on Medical and Aromatic Plants*, 5: 72-81.
- Zhang, X., Li, Q., Xu, Z. and Dou, J. (2020). Mass spectrometry-based metabolomics in health and medical science: A systematic review. RSC Advances, 10: 3092-3104.
- 8. Shaharuddin, N. H., Ismail, N. H., Manshoor, N., Salim, F. and Ahmad, R. (2016). Chemical profiling and identification of alkaloids and flavonoids in *Uncaria lanosa* var. *ferrea* via UHPLC-Orbitrap MS. *Malaysian Journal of Analytical Sciences*, 20(2): 318-323.

- 9. Salim, F., Ismail, N. H., Awang, K. and Ahmad, R. (2016). Rauniticine-*allo*-oxindole B and rauniticinic-*allo* acid B, new heteroyohimbine-type oxindole alkaloids from the stems of Malaysian *Uncaria* longiflora var. pteropoda. *Molecules*, **16**(8): 6541-6548.
- 10. Salim, F., Yunus, Y. M., Anouar, E. H., Awang, K., Langat, M. K., Cordell, G. A., and Ahmad, R.
- (2019). Absolute configuration of alkaloids from *Uncaria longiflora* var. *pteropoda* through experimental and theoretical approaches. *Journal of Natural Products*, 82: 2933-2940.
- 11. Salim, F. and Ahmad, R. (2010). Isopteropodic acid from Malaysian *Uncaria longiflora* var. *pteropoda*. *World Applied Science Journal*, 10(11): 1334-1337.