

## CHEMICAL CHARACTERIZATION OF *Berberis vulgaris* L. var. *asperma* EDIBLE BERRIES

(Pencirian Kimia Buah Beri Boleh di Makan *Berberis vulgaris* L. var. *asperma*)

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### Abstract

*Berberis vulgaris* L. var. *asperma* edible fruits have been found to have activity against several microbes and to be cytotoxic in certain immortalized cancer cell lines. In this study, the berries were analyzed to find the possible metabolites that could be the curative agents. Evaluations using an LC-UV/MS technique established the presence of chlorogenic acid, rutin, and berberine in ethanolic extracts of *B. vulgaris* berries. The ethanolic sample was found to have 47.1 mg/kg of 3-(3,4-dihydroxycinnamoyl) quinic acid, 0.02 mg/kg of rutin and 0.36 mg/kg of berberine. The free radical scavenging activity of the mixture of polyphenolics exhibited an IC<sub>50</sub> at 34.48 µg/mL. An assessment of zinc and iron concentration using atomic absorbance spectrophotometry (AAS) was determined to be 45.3 and 198.1 mg/kg, respectively. The investigation of the volatile constituents through solid phase microextraction (SPME) and by dichloromethane extraction was explored using a gas chromatograph equipped with a mass spectrometer (GC-EI-MS). SPME analyses showed the presence of palmitic acid (C16:0), vaccenic acid (C18:1 Δ 11 trans) and stearic acid (C18:0). Three saturated fatty acids, SFAs (C14:0, C16:0, and C18:0), two methyl esters of the SFAs (C14:0 and C16:0), and one polyunsaturated fatty acid, PUFA (C18:2 Δ 9,12 cis) were detected in the dichloromethane extracts of the berries.

**Keywords:** atomic absorption spectroscopy, *Berberis vulgaris*, mass spectrometry

### Abstrak

Buah yang boleh dimakan *Berberis vulgaris* L. var. *asperma* telah diketahui mempunyai aktiviti melawan beberapa jenis mikroba and menjadi sitotoksik di dalam sesetengah titisan sel kekal. Dalam kajian ini, buah beri dianalisis untuk mencari kebarangkalian metabolit yang boleh digunakan sebagai agen penawar. Penilaian menggunakan teknik LC-UV/MS dibangunkan untuk mengesan kehadiran asid klorogenik, rutin dan berberin di dalam ekstrak beri *B. vulgaris*. Sampel etanolik ditemui mengandungi 47.1 mg/kg 3- (3,4 – dihidroksisinasamoyl) asid quinik, 0.02 mg/kg rutin dan 0.36 mg/kg berberin. Pemerangkapan aktiviti radikal bebas bagi campuran polifenolik wujud pada IC<sub>50</sub> 34.48 µg/mL. Penilaian kepekatan zink dan besi menggunakan spektrofotometri serapan atom (AAS) telah ditentukan dengan masing-masing ialah 45.3 dan 198.1 mg/kg. Penyiasatan terhadap jujukan sebatian merupa dilakukan menggunakan pengekstrakan mikro fasa pepejal (SPME) dan pengekstrakan diklorometana sebelum di analisis menggunakan kromatograf gas dilengkapi spektrometer jisim (GC-EI-MS). Analisis SPME menunjukkan kehadiran asid palmitik (C16:0), asid vasetik (C18:1 Δ 11 trans) dan asid stearik (C18:0). Tiga asid lemak tepu, SFAs (C14:0, C16:0, dan C18:0), dua metal ester SFAs (C14:0 and C16:0), and satu asid lemak politaktepu, PUFA (C18:2 Δ 9,12 cis) telah dikesan di dalam ekstrak diklorometana buah beri.

**Kata kunci:** spektroskopi serapan atom, *Berberis vulgaris*, spektrometri jisim

### Introduction

*Berberis vulgaris* is a member of the Berberidaceae family and a common garden bush found in Europe and the British Isles. Barberry, as it is commonly known, has been used by herbalists since ancient times with its usage dating back to the Egyptians. Alkaloids and flavonoids have been reported in the root, stem, leaves and fruit of this bush. These two compound classes have been found to exhibit anti-diarrheal, anti-microbial, anti-inflammatory, anti-allergic, and anti-cancer effects. The edible fruit of *B. vulgaris* has been used in Asia and Europe especially in Iran as traditional medicine for various diseases which include malaria, dysentery and gall stones. Its antihistaminic and anticholinergic activity was found in the aqueous extracts of barberry on isolated guinea pig ileum [1]. Antioxidant activities of ethanolic extracts of roots, twigs, and leaves of the barberry plant were correlated with phenols and flavonols present in the plant tissue [2]. The fruit has been linked to diseases of the kidneys, urinary or gastrointestinal tract, liver, bronchial, and as a stimulant for the circulatory system [3].

In this work, ethanolic extracts of the barberry fruit were analyzed using a radical scavenging assay, high-performance liquid chromatography (HPLC), liquid chromatography-mass spectrometry (LC-ESI-MS), gas chromatography-mass spectrometry (GC-EI-MS), solid-phase microextraction (SPME) and dichloromethane extracts, and atomic absorption spectrometry (AAS). To the best of our knowledge, this is the first reported study using this methodology of chemical analyses on the extracts of *B. vulgaris* berries.

### Materials and Methods

Fresh samples of *B. vulgaris* fruits were harvested from Mashhad, Khorasan Province, Iran and identified at the Mashhad Medical University of Sciences, Iran. The berries were lyophilized and extracted with ethanol for 10 hours, filtered (22 µm pore size), and concentrated in vacuo. The residue was reconstituted with ethanol (60 µg/µL) and was passed through a 0.45 µm cellulose acetate membrane before further analysis.

#### Free radical scavenging assay

The radical scavenging activity of *B. vulgaris* ethanolic fruit extracts was measured by 2,2-Diphenyl-1-picrylhydrazyl or DPPH (Sigma-Aldrich, Singapore) utilizing a modified version of the method by Sikder et al. [4]. An amount 10 mL of 60 mM of DPPH was added to each trial where the extracts were varied from 5 µL to 125 µL. The resulting reaction mixtures of 30.00 to 740.74 µL/mL were incubated at ambient temperatures in the dark for 30 minutes. A control sample which contained only DPPH solution and no fruit extract was also made. After incubation, changes in absorbance at 517 nm were analyzed using a Hitachi 2900 double beam UV-Visible spectrophotometer. Nonlinear regression and statistical analyses were done using GraphPad Prism 7.01 (GraphPad Software, Inc.) to extrapolate the half maximal inhibitory concentration, IC<sub>50</sub> (50% reduction of absorbance as compared to the control).

#### RP-HPLC analyses

Fruit extracts (20 µL) were analyzed using an Agilent Technologies 1200 Series quaternary pump system with a variable wavelength detector and a 20-µL sample injector. Analytes were separated on a 250 mm × 4 mm, 5-µm particle RESTEK Pinnacle II ODS column (Supelco). Resolved compound peaks were identified by direct comparison of their retention times with reference standards. The columns were maintained at room temperature.

Optimization was conducted using a method developed by Hasler et al. [5] and Olszewska [6]. Solvent system consisted of 5% formic acid in distilled water (FA/H<sub>2</sub>O) and acetonitrile (ACN) with a gradient of: 0-2 minutes at 95% (FA/H<sub>2</sub>O) and 5% ACN; 5-7 minutes at 90% (FA/H<sub>2</sub>O) and 10% ACN; 7-10 minutes at 90-85 % (FA/H<sub>2</sub>O) and 10-15% ACN; 10-13 minutes at 85-80% (FA/H<sub>2</sub>O) and 15-20% ACN; 13-23 minutes 80% (FA/H<sub>2</sub>O) and 20% ACN; 23-25 minutes at 80-70% (FA/H<sub>2</sub>O) and 20-30% ACN; 25-28 minutes at 70% (FA/H<sub>2</sub>O) and 30% ACN; 30-32 minutes at 50% (FA/H<sub>2</sub>O) and 50% ACN; 32-45 minutes at 50-95% (FA/H<sub>2</sub>O) and 50-5% ACN. Flow rate was 1.00 mL/min, while the UV detector was set at 350 nm for all the trials. Standard stock solutions initially from 500 ppm of berberine (Sigma-Aldrich, Singapore), rutin (Sigma-Aldrich, Singapore), and chlorogenic acid (Sigma-Aldrich, Singapore) were freshly prepared in methanol and the goodness of fit generated from the absorbance per concentration on scatter charts confirmed whether the linear regression analysis ( $R^2 > 0.97$ ) and external calibration were acceptable.

### LC-ESI-MS analyses

Fruit extracts were analyzed using LC-MS system equipped with DIONEX Ultimate 3000 HPLC & Bruker micrOTOF-Q II MS with a C<sub>18</sub> column (3 mm, 120 Å, 2.1 x 150 mm). Electrospray ionization positive mode was used and acquisition was at 50 m/z to 3000 m/z. The solvent gradient for RP-HPLC was used, with the last segment of the gradient extended for five minutes to account for the internal calibration (sodium formate) that was run simultaneously with the trials. Other parameters were as follows: capillary at 4500 V, end plate offset at -500 V, collision cell RF 500.0 VPP, nebulizer at 2.0 bar, dry heater at 180 °C, dry gas at 8.0 L/min. Mass spectra were calibrated internally during the trials and the background subtracted during data analysis.

### SPME-GC-EI-MS analyses

Approximately 7 grams of lyophilized fruit extracts were placed in an Erlenmeyer flask covered with a septum. Samples for GC-MS analysis was collected using a polydimethylsiloxane (PDMS) fiber while heating the set-up at 40 °C for 30 minutes. The fiber was desorbed and analyzed manually in a Perkin–Elmer Clarus 500 gas chromatograph - mass spectrometer. Sampling rate was 1.56250 points/second. Temperature was set at 40 °C for 5 minutes then increased by 4 °C until it reached 230 °C where it was held for 5 minutes. Flow rate of the carrier gas was set at 2.0 mL/min while ionization was carried out using 70 eV (EI), and the masses were scanned from 50 m/z to 500 m/z.

### GC-EI-MS analyses of DCM extracts

The freeze-dried *B. vulgaris* berries (9.37 g) were ground in an osterizer, soaked in CH<sub>2</sub>Cl<sub>2</sub> (DCM) for one day, filtered, and concentrated over nitrogen gas to achieve the resultant crude residue (5.25 g) which was then redissolved in 1 mL of DCM. An Agilent GC MS 7890B, equipped with a HP-5ms (5% phenyl methylsiloxane) Ultra Inert column (30 m x 250 mm x 0.25 mm) with ultra-pure helium gas as the mobile phase, was used for the analysis of volatile constituents. The flow rate of helium gas was at 1 mL/min, pressure maintained at 8.2 psi, with an average velocity of 36.62 cm/sec and holdup time of 1.37 minutes. The split inlet, at 20:1 ratio, was held at 250 °C, at 8.2 psi, total flow of 24 mL/min, with a septum purge flow rate of 3 mL/min. The temperature for the injector was held at 250 °C and optimization of the temperature program was started at 50 °C with a programmed linear ramp of 5 °C/min until 220 °C.

Compound identification was done using the NIST library 2.0 and peak areas were processed from the resultant total ion chromatograms. The resultant data was confirmed by the comparison of the compounds according to their elution order with their relative retention indices on a non-polar stationary phase. The retention indices were computed for all of the volatile constituents utilizing a homologous series of n-alkanes.

### AAS analysis

Approximately 1 g of lyophilized fruit extract was pulverized and heated in a furnace at 200 °C to 250 °C for 30 minutes. Then, the sample was reduced to ashes by heating at 450 °C for 4 hours. After this, 2 mL of 5 M HNO<sub>3</sub> was added and the resulting mixture was evaporated to dryness in a sand bath. The sample was further dried at 400 °C for 15 minutes, then the ashes were dissolved in 5 mL of 20% HCl followed by dilution to 25 mL. The zinc and iron content of the sample were analyzed using a Shimadzu AA-6300 atomic absorption spectrophotometer.

## Results and Discussion

### Free radical scavenging activity

Through curve fitting or nonlinear regression of the data points of the varying volumes of barberry extract, the IC<sub>50</sub> was found to be 34.48 µg/mL. The reaction mixture with 149.63 ppm of barberry extract caused the maximum decrease in absorbance and additional increments of the ethanolic extracts did not exhibit any significant variance ( $p > 0.05$ ) at 95% CI of discrepancy. The extent of decrease in absorbance from the control correlates with the free radical scavenging potential of the antioxidant present in the plant extract [7].

### Identification of alkaloids and flavonoids

Mass spectral analysis of *B. vulgaris* ethanolic fruit extracts resulted in the identification of three compounds (Figure 1). At a retention time of 9.9-10.2 minutes and a molecular weight of 355.1 m/z, the chemical formula C<sub>16</sub>H<sub>19</sub>O<sub>9</sub> with -0.1 err [mDa], -0.2 err [ppm], and 2.9 mSigma correlated with [M]<sup>+1</sup> of chlorogenic acid or

3-(3,4- dihydroxycinnamoyl) quinic acid. Rutin,  $[M+H]^+$ , was found at a retention time of 14.2 - 14.9 minutes at 611.2 m/z, with the chemical formula  $C_{27}H_{31}O_{16}$  and 1.4 err [mDa], 2.2 err [ppm] and mSigma 4.5. The  $[M+H]^+$  of berberine,  $C_{20}H_{18}NO_4$ , was found at 17.6 - 17.8 minutes.

#### Quantitative analysis of chlorogenic acid, berberine and rutin

From the chromatograms of the ethanolic extract and comparison of the retention times, it was found that chlorogenic acid (polyphenol), rutin (flavonoid) and berberine (alkaloid) were present. Analysis using chlorogenic acid, rutin, and berberine standards gave average retention times of 10.39, 18.58, and 33.81 minutes, respectively. The barberry extracts were found to contain 47.1 mg/kg of chlorogenic acid, 0.02 mg/kg of rutin and 0.36 mg/kg of berberine.

Chlorogenic acid has been found to be an activator of the Nrf2/antioxidant-response element pathway, which was initiated during oxidative stress [8]. The extensive range of therapeutic effects, such as antimutagenic, antiviral [9], anticarcinogenic [7], and free radical scavenging activity [10] has led many researchers to believe that this compound can have potential pharmaceutical applications. Rutin (vitamin P or rutoside) is considered a high value nutraceutical whose pharmacological action has been reported to benefit the central nervous system through its neuroprotective effect on brain ischemia by the upregulation of 'endogenous antioxidant defense enzymes' [11]. A significant attenuation in HL-60 cells in a murine model using a dose of 120 mg/kg of rutin displayed a high antileukemia capacity [12]. Berberine was found to inhibit gram-positive bacteria such as *Staphylococcus aureus* (X) with a MIC value of 250 mg/L [13]. Inhibition of mutant keratinocytes by berberine at  $IC_{50} = 30 \mu\text{mol/L}$  [14] and in HeLa cells at 100 and 150 mg/L [15] were observed.

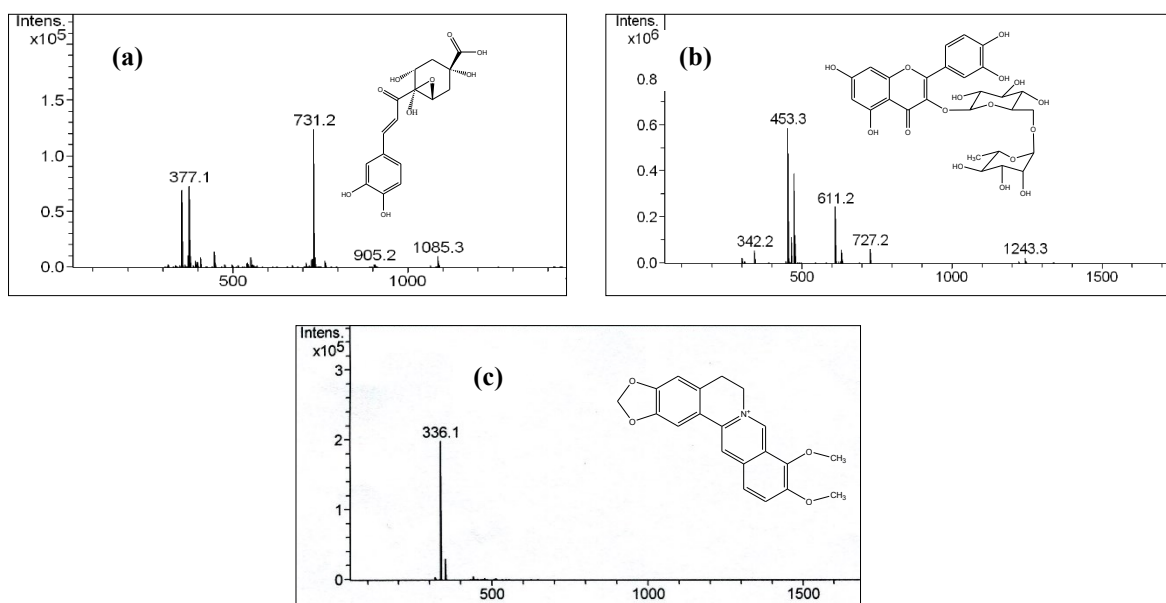


Figure 1. Mass spectra and structures of compounds (a) 1, (b) 2, and (c) 3 found in the LC-ESI-MS analyses

#### Concentration of zinc and iron

The barberry samples contained 45.3 and 198.1 mg/kg of zinc and iron respectively. *Streptococcus pyogenes*, *Staphylococcus aureus* and *Pseudomonas aeruginosa* were found to have physical damage on the bacterial cell membranes due the adhesion of zinc oxide rod structures on the sheaths [16]. Inorganic metal oxides such as iron(II) oxide can also act as operative antiseptics, owing to their innocuous profile, chemical stability and bioactivity against certain microbes. Composite nanoparticle structures oxides of Zn/Fe exhibited significant bacteriostatic activity on *Staphylococcus aureus* and minimally against *Escherichia coli* [17].

### SPME-GC-EI-MS Analysis

Palmitic acid (256.0128 m/z,  $[M]^+$ ) at 45.10 minutes was observed and the spectra showed the fragment  $[(CH_2)_2CO_2H]^+$  with m/z = 72.9138. Fragments at 59.9186, 128.9987, 157.0768, 213.0133, and 227.0895 m/z also exhibited ions with the formula  $[(CH_2)_nCO_2H]^+$ . Vaccenic acid was the most probable compound at 50.29 minutes which manifested a molecular ion peak at 282.1020 m/z and a base peak at 54.930 m/z which is presumed to be  $[C_4H_7]^+$ . The peak at 50.77 minutes gave a fragmentation pattern attributed to stearic acid 284.1314 m/z  $[M]^+$ ; the base peak was also at 72.9138 m/z and exhibited a similar fragmentation pattern to that of palmitic acid. Olefinic fragments at approximately m/z: 69, 83, and 97 were also found in the three mass spectra (Figure 2).

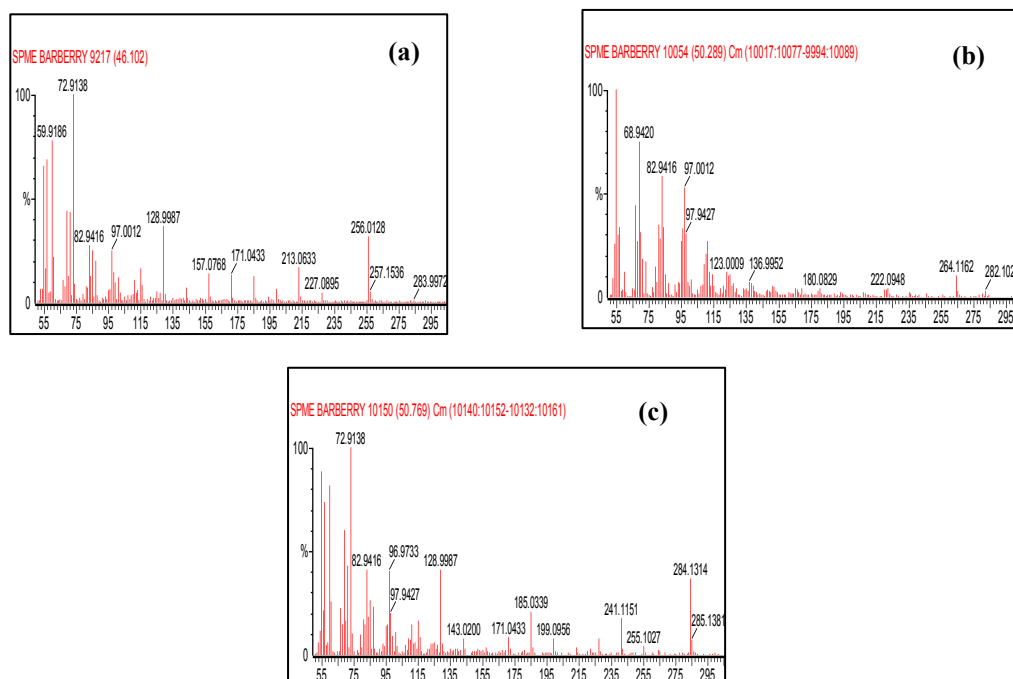


Figure 2. Mass spectra of (a) palmitic acid, (b) vaccenic acid, (c) stearic acid in the SPME-GC-EI-MS analyses

### GC-EI-MS analyses of DCM extracts

GC-MS analyses of the dichloromethane extracts of *B. vulgaris* edible berries led to the identification of six constituents. The identified components of the low boiling point compounds of the crude extract were substantiated by retention index (RI) and structural class through the NIST library. The results are listed in Table 1 according to their elution order on a HP-5MS column. The sample consisted primarily of three SFAs (C14:0, C16:0, and C18:0), two methyl esters of the SFAs of C14:0 and C16:0, and one PUFA (C18:2  $\Delta$  9,12 cis).

Antimicrobial function in human milk, skin and mucosal membranes have been linked to the presence of fatty acid in these food products which may have the capacity to be useful as microbicides against human pathogens that occur on the skin and mucosa [18, 19, 20]. Various concentrations of saturated fatty acids (SFAs) capric acid (C10:0), lauric acid (C12:0), myristic acid (C14:0), palmitoleic acid (C16:1), monolaurin (C10:0) monolaurin (C12:0) and monopalmitolein (C16:0) caused significant reduction in *H. pylori* cells [21]. The natural trans-fat vaccenic acid is the natural precursor of the polyunsaturated fatty acid (PUFAs) linoleic acid (C18:2  $\Delta$  9,12 cis) and has been reported to have hypo-triglyceridemic action in JCR:LA-cp rats [22] and have anti-proliferative effects on the immortalized cell line MCF-7 (breast adenocarcinoma) by inhibiting the expression of Bcl-2 and procaspase-9 [23].

Table 1. The volatile constituents of the *B. vulgaris* DCM extracts

Peak Number	RT (minutes)	Compound <sup>b</sup>	RI <sup>a</sup>	Functionality
1	14.75	tetradecanoic acid	1766	saturated fatty acid
2	17.55	hexadecanoic acid methyl ester	2026	fatty acid methyl ester
3	20.08	n-hexadecanoic acid	1962	saturated fatty acid
4	20.66	9,12- octadecanoic acid (z, z)-	2068	polyunsaturated fatty acid
5	21.21	octadecanoic acid	2082	saturated fatty acid
6	21.33	tetracosanoic acid, methyl ester	2433	fatty acid methyl ester

<sup>a</sup> Retention Index (HP-5ms column), <sup>b</sup> Compounds listed in order of elution from a HP-5MS column

### Conclusion

The bioactivity of *B. vulgaris*, which has been found to exhibit anti-microbial, anti-inflammatory, and anti-cancer effects, can be linked to the chemical constituents found inherently in the fruit. The edible berry was established to contain significant amounts of chlorogenic acid (47.1 mg/kg), rutin (0.02 mg/kg) and berberine (0.36 mg/kg) which have been observed to have medical applications and could be responsible for the observed high antioxidant potential in the DPPH assay ( $IC_{50} = 34.48 \mu\text{g/mL}$  crude extract). The bacteriostatic potential of these constituents could be linked to the ability of these entities to penetrate cell membranes, thereby affecting the integrity of the microbes, and to inhibit quorum-sensing. A substantial amount of zinc, iron, and medium to long chain free fatty acids were also detected in the samples which have been reported to induce the degradation of pathogens and malignant neoplasms. The pharmacognosy involved could be correlated to the translational capacity of these constituents to induce caspase-dependent apoptosis, quenching of reactive oxygen species, regulation of phase I or phase II enzymes, and the expression of anti-inflammatory agents.

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